### **Databases for Gene Expression analysis**

Perform a THOROUGH in silico analysis before doing benchwork

What is the difference between databases and papers?

#### For your pet gene:

determine all names using genecards
how many known exonic SNPs are there
find two commercial antisera for the protein
get the complete sequence for your pet gene using the UCSC database and
compare the exon intron-organisation with the one predicted by exonscan

### There are multiple databases for expression analysis

Abbroviation Database

### http://www.science.co.il/biomedical/RNA-Databases.asp



	5S ribosomal RNA database							
ASTRA	Alternative Splicing and TRanscription Archives							
AAARS	Aminoacyl-tRNA synthetases database							
	Codon Usage Database							
	Comparative RNA Web Site							
Mamit-tRNA	Compilation of mammalian mitochondrial tRNA genes							
	Compilation of tRNA sequences and sequences of tRNA genes							
PolyA_DB	Database for mammalian mRNA polyadenylation							
snoRNABase	Database of Human H/ACA and C/D Box snoRNAs							
BPS	Database of RNA Base pair Structures							
Recode	Database of translational recoding events							
	DNA and RNA Molecular Weights and Conversions							
	European Ribosomal RNA Database							
FSDB	Frameshift Database							
	Free Energy and Enthalpy Tables for RNA Folding							
fRNAdb	Functional RNA Database							
sRNAMap	Genomic maps for small non-coding RNAs, their regulators and their targets in microbia genomes $$							
GtRDB	Genomic tRNA Database							
	Greengenes: 16S rDNA gene database and tools							
H-InvDB	H-Invitational database of human genes and transcripts							
RNAdb	Mammalian noncoding RNA database							
	Methylation Guide snoRNA Database							
miRBase	microRNA Sequence Database							
	Modomics: a database of RNA modification pathways							
ncRNA	Noncoding RNA database							
RNaseP	Ribonuclease P Database							
RDP	Ribosomal Database Project of 16S rRNA sequences							
	Ribosomal RNA Mutation Database							
RAG	RNA As Graphs Web resource							
Rfam	RNA families database							
	RNA Modification Database							
STRAND	RNA secondary STRucture and statistical ANalysis Database							
	Silva rRNA database							
	Small Subunit rRNA Modification Database							
SCOR	Structural Classification of RNA							
tmRDB	tmRNA Database							
	tmRNA Website							
	Viral RNA Structure Database							
	Yeast Small Nucleolar RNA (snoRNA) Database							

### Five useful sites

Genecards, <a href="http://www.genecards.org/">http://www.genecards.org/</a>
Fast overview over a gene, link to reagents

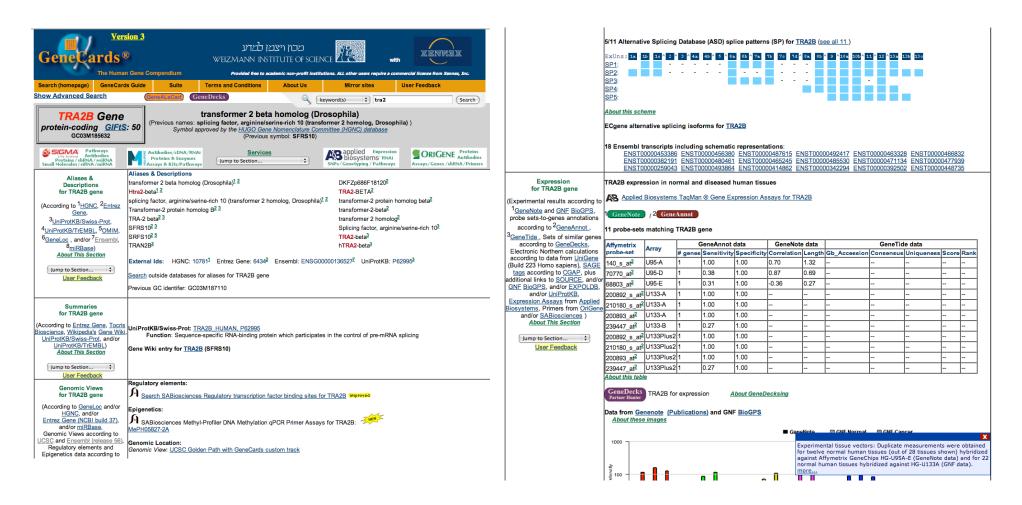
UCSC genome browser, <a href="http://genome.ucsc.edu/">http://genome.ucsc.edu/</a> Most comprehensive (and best to use) database

Fast DB, <a href="http://www.fast-db.com/fastdb2/frame.html">http://www.fast-db.com/fastdb2/frame.html</a>
Specialized in splicing, very easy to use,
<a href="mailto:Log in: Stefan@stamms-lab.net">Log in: Stefan@stamms-lab.net</a>, PW: OUSMqllerX

Gil Ast lab page, <a href="http://www.tau.ac.il/~gilast/">http://www.tau.ac.il/~gilast/</a> Links to splicing analysis tools

Christopher Burge lab, http://genes.mit.edu/burgelab/software.html Links to splicing analysis tools

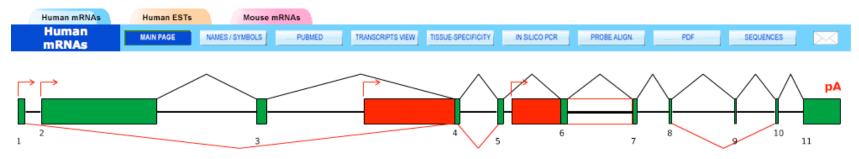
# Genecards



Fast, comprehensive overview for a given gene (expression, names, publications...)

# **FAST-DB**

## splicing factor, arginine/serine-rich 10 (transformer 2 homolog, Drosophila) (ID 15379)



[ 21284 bp ]

[ chromosome (strand): 3(-):187117238-187138521 ]

[ 11 EXON(S) ]

[ legend ]

TRA	ANSCRIPTION INITIATION & FIRST	EXON(S)	
	FIRST EXON(S)	Number of evidence(s)	
	EXON 1	9	
	EXON 2	1	
	EXON 4	1	

>exon 5 163 bp (12406-12568)
GTCTAGATCCAGAAGAAGCTCCCGAAGGCATTATACCC
GGTCACGGTCTCGCTCCCGCTCCCATAGACGATCACGT
AGCAGGTCTTACAGTCGAGATTATCGTAGACGGCACAG
CCACAGCCATTCTCCCATGTCTACTCGCAGGCGTCATG
TTGGGAATCGG

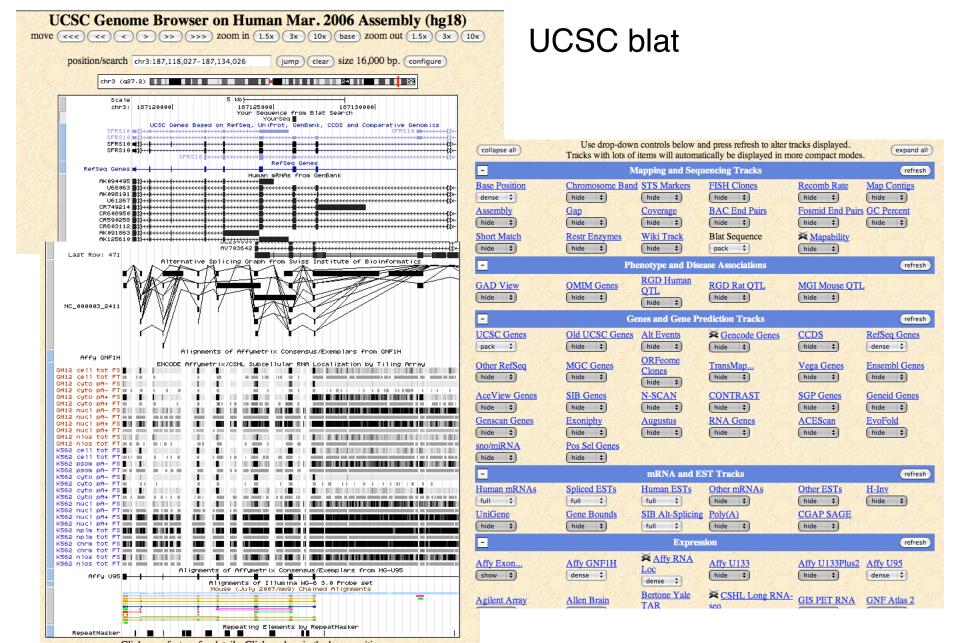
http://www.fast-db.com

User-friendly

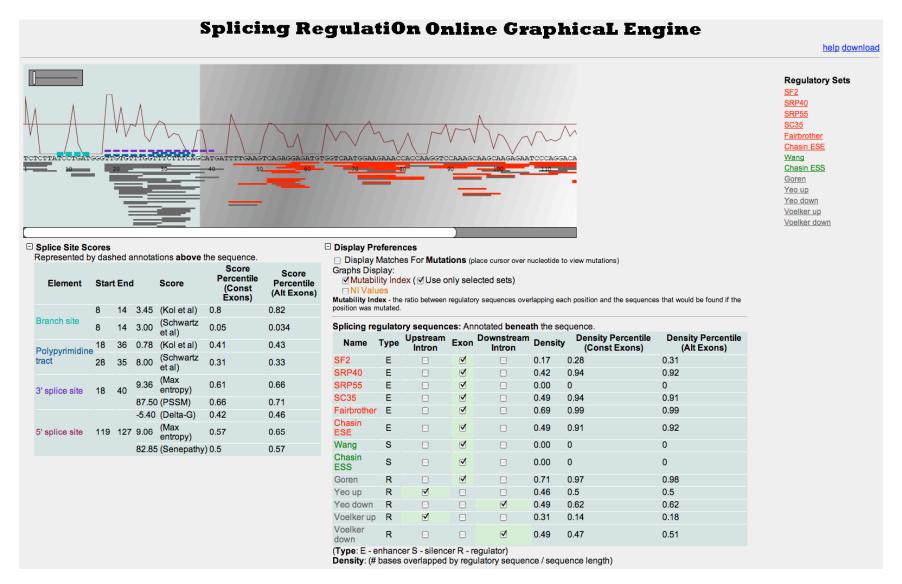
Stefan@stamms-lab.net

PW: OUSMqllerX

### UCSC genome browser



# Gil Ast homepage



#### **Christopher Burge lab**

#### **EXONSCAN**

### ExonScan results page

locus: HUMATPGG length: 12900 cutoff: 192 Predicted exons:

Begin	-	End	(:	3'ss	5'ss	ESE	ESS	GGG	total)
35	_	178	(	92	37	102	6	39	276)
294	-	353	(	50	96	12	0	51	209)
925	-	1128	(	97	52	3	13	39	204)
2631	-	2744	(	101	85	19	-1	9	213)
2948	-	3029	(	78	58	75	10	6	227)
4371	-	4569	(	90	63	36	-3	27	213)
5066	-	5200	(	79	91	9	-2	21	198)
5715	-	5907	(	109	85	25	5	9	233)
7751	-	7887	(	117	74	15	-2	15	219)
7973	-	8123	(	97	80	34	3	21	235)
8487	-	8641	(	89	85	66	0	30	270)
9615	-	9856	(	116	43	27	-4	39	221)
10380	-	10525	(	132	72	33	-3	6	240)
12450	-	12551	(	95	85	25	5	24	234)
12636	-	12727	(	104	60	16	14	39	233)

#### Key:

- locus: gene name (taken from line beginning with > if present)
- · length: number of bases in given sequence
- · cutoff: minimum total exon score to be considered for prediction
- · Begin: index (starting at one) of first base in predicted exon
- End: index (starting at one) of last base in predicted exon
- 3'ss: maximum entropy score of 3' splice site (beginning of exon)
- 5'ss: maximum entropy score of 5' splice site (end of exon)
- ESE: sum of scores for all RESCUE-ESEs in predicted exon
- . ESS: sum of scores for all FAS-hex3 ESSs in predicted exon
- GGG: sum of scores for all GGGs in certain intronic regions flanking predicted exon
- · total: sum of all other scores

All scores are in tenths of bits.

Return to ExonScan Web Server

#### SUMMARY AND OUTLOOK

Lots of information available about gene expression that can be visualized using browsers

Problem with databases is their upkeep, UCSC and ENSEMBLE up to date for nucleic acids

Multiple analysis tools are available on the web

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